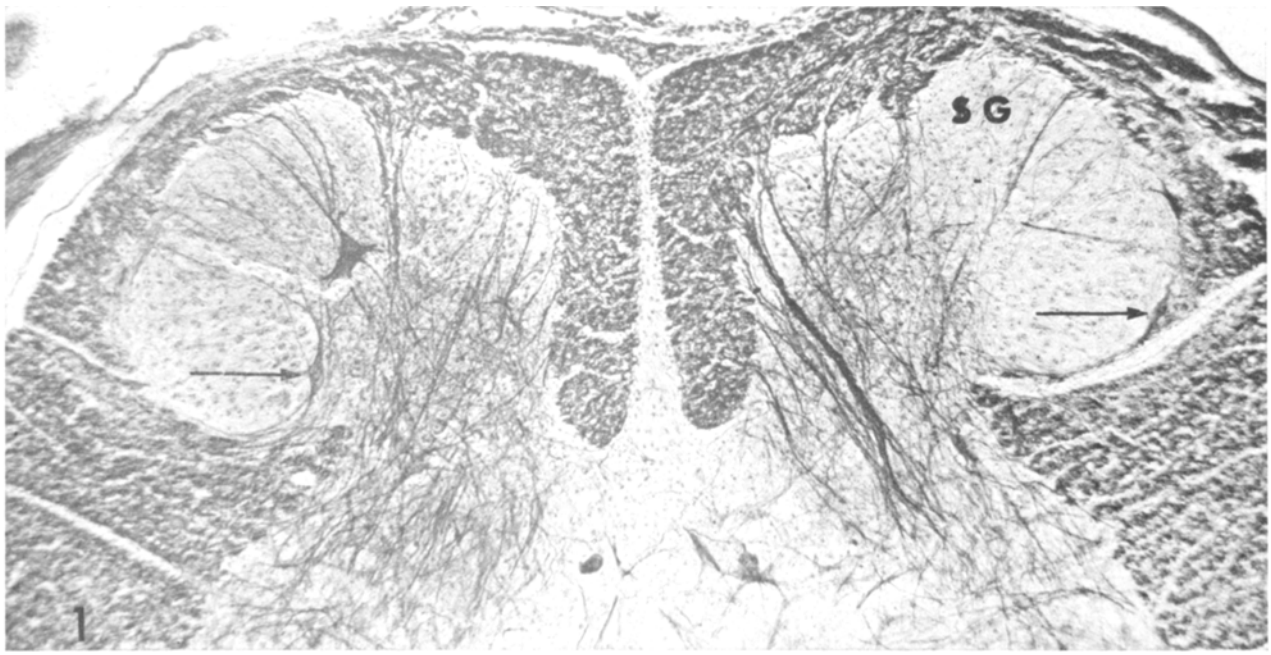
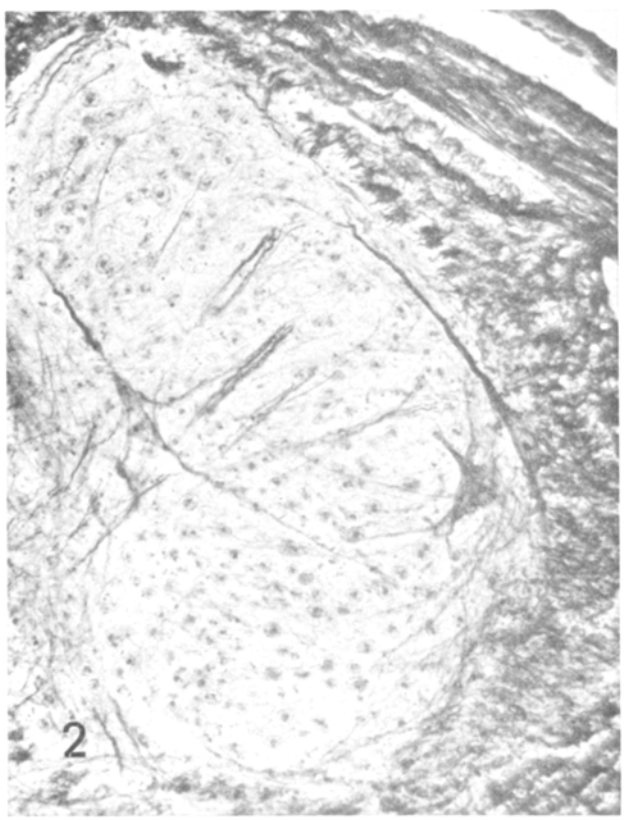
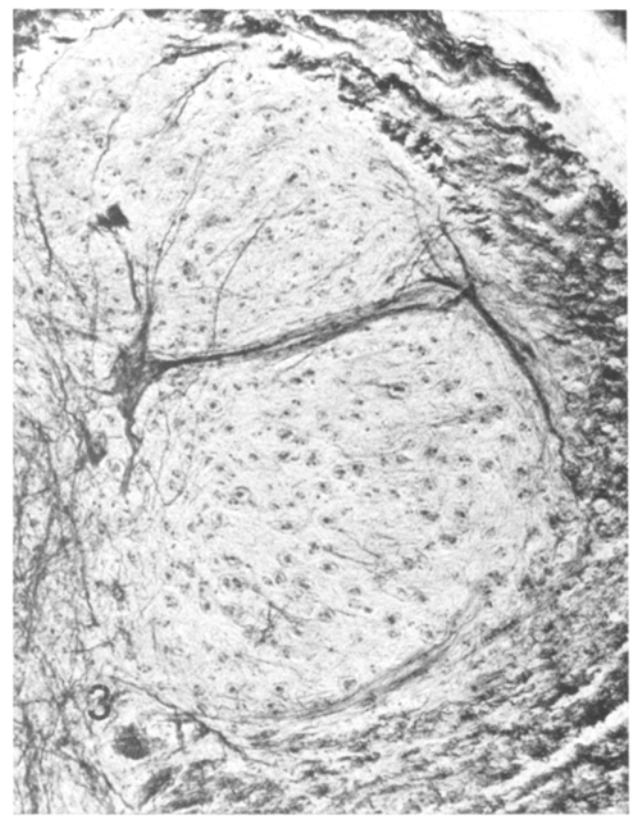
Large Neurons in the Substantia Gelatinosa of Rolando

The dorsal horn of the vertebrate spinal cord has been examined extensively by investigators using both light and electron microscopic techniques (CAJAL[1], KEENAN[2], REXED[3], SCHEIBEL and SCHEIBEL[4], PETRAS[5], SZENTAGOTHAI[6] and RALSTON[7]). These studies have established that an area of the dorsal horn, the substantia gelatinosa of Rolando is composed of small Nissl free neurons and fine nerve fibers. This report will describe a further cell

[1] R. Y. CAJAL, *Histoire du Système Nerveux* (C.S.I.C., Madrid 1952), p. 330.
[2] E. KEENAN, Koningl. Akad. Wetenschap. Amsterdam *32*, 466 (1929).
[3] B. REXED, Progr. Brain Res. *2*, 58 (1964).
[4] M. SCHEIBEL and A. SCHEIBEL, Brain Res. *9*, 32 (1968).
[5] J. M. PETRAS, Experientia *24*, 1045 (1968).
[6] J. SZENTAGOTHAI, J. comp. Neurol. *122*, 219 (1964).
[7] H. J. RALSTON III, Z. Zellforsch. *67*, 1 (1965).







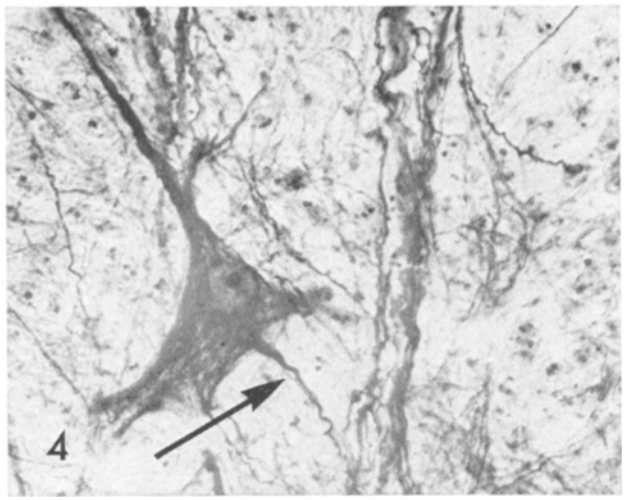


Fig. 1–4. Silver stained transverse sections from the spinal cords of chick embryos incubated for 18 days. The arrows in Figure 1 point to marginal cells while the arrow in Figure 4 indicates an axon extending towards the dorsal white matter.

type found in the substantia gelatinosa of the embryonic and adult avian spinal cord.

Materials and methods. Chick embryos incubated for 12–20 days and post hatched chicks up to 5 weeks of age were sacrificed and the spinal cords fixed in Carnoys B fluid for Nissl staining or in De Castros fixative for block Cajal staining. The cords were embedded in paraplast, sectioned serially at 15 μ and mounted on glass slides.

Results. In Nissl and silver stained sections of embryonic spinal cord the substantia gelatinosa forms a well defined band extending from the medial side of the dorsal cell column, around the apex and down the lateral side, ending near the base of the dorsal horn. It is composed primarily of small Nissl free neurons, however, an occasional large multipolar neuron, average perikaryal diameter 35–45 μ is also present (Figure 1). These large neurons have spherical nuclei with 1 or 2 darkly stained nucleoli and their cytoplasm contains large floccular Nissl granules. The large multipolar neurons, usually 1 per section occasionally 2 as shown in Figure 2, are located in fairly fixed positions. When only one is present, the cell body is located near the apex of the dorsal cell column (Figures 1–3). If a second cell is present it is usually found close to the lateral boundary of the substantia (Figure 2). The processes of these large multipolar neurons can be followed in various directions with the dendrites oriented towards the dorso-lateral aspect of the cord and the axons (arrow in Figure 4), extending into the ventro-medial area of the dorsal white matter.

Discussion. Although this report has dealt primarily with the embryonic spinal cord it must be stressed that the large multipolar neurons were also found in the substantia of 5-week-old post hatched chicks. The cells in question however attain their maximum size and are largest, relative to their surroundings, during the last 5 days of incubation. The maturation of these large neurons in the substantia, coincides with the onset of repetitive somatic movements of the chick prior to hatching and it is interesting to speculate that these cells may be part of a transient system concerned with the hatching behavior.

Résumé. On a trouvé chez le poulet, dans la substance gélatineuse de la moëlle épinière en voie de maturation ou adulte, de gros neurones multipolaires d'un diamètre moyen de 35–45 μ. Ces neurones ont des noyaux sphériques contenant un ou deux nucléoles fortement contrastés. Dans leur cytoplasme se trouvent des granules de Nissl, gros et à contour floconneux.

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Incorporation of ^{35}S-Labelled L-Cysteine in the Ependyma of the Rat's Subcommissural Organ and Choroid Plexus

Conspicuous incorporation of ^{35}S-labelled dl-cysteine in the ependyma of the subcommissural recess has previously been elicited by autoradiographs[1]. Observations have been made to the effect that changes in the function of the thyroid gland have an influence on this incorporation[2].

The problematic function of the subcommissural organ (SCO) has been a constant object of investigation. Endeavours have been made to find new methods of research for clarification of its physiological significance. Particular efforts have been exerted on the search for a morphological method to measure the activity of the SCO, in addition to the previously predominantly employed quantification of the selectively stainable 'secretion' and to karyometric measurement. The incorporation of cysteine has been used as indicator for the activity of the hypothalamic-hypophyseal neurosecretory system. Similarly, this incorporation may be thought to reflect also the secretory activity of the SCO. Closer analysis of the phenomenon was therefore considered appropriate. In order to obtain a suitable reference basis, the incorporation in the ependyma of the choroid plexus of the third ventricle was also studied in addition to that taking place in the ependyma of the SCO.

Material and methods. Altogether 31 adult male albino rats of 225 g average body weight were used. The animals received standard pellet diet and tap water ad libitum. All the rats were kept under identical conditions. Each animal was given, by i.p. injection, ^{35}S-labelled L-cysteine ('L-Cysteine-S 35 hydrochloride', The Radiochemical Centre, Amersham, Bucks., England) at an average dose of 150 μCi. The injections were all administered on one day, about 09.00 h. The animals were divided into 11 groups, which were sacrificed at the following times after injection: 10, 30 and 45 min, 1, $1^1/_2$, 2, 4 and 6 h, 1, 2 and 3 days. There were 3 rats in every group but those sacrificed after 2 and 3 days, which had 2 members each. The rats were killed by rapid decapitation without anaesthesia.

The brains were embedded in paraffin after fixing in Bouin's fluid. Serial sections at 7 μ were made sagittally;

[1] J. C. Sloper, D. J. Arnott and B. C. King, J. Endocrin. *20*, 9 (1960).
[2] S. Talanti and V. Pasanen, Life Sci. *7*, 1245 (1968).